



Hong Kong Institute of Medical Laboratory Sciences

Quality Assurance Programme

May Survey (2024)

Anatomical Pathology

HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES

QUALITY ASSURANCE PROGRAMME

Anatomical Pathology

INSTRUCTIONS

1. All tests should be performed as soon as possible after the receipt of the specimens.
2. Unless otherwise specified, all survey material had been fixed in 10% buffered formalin and paraffin embedded.
3. The survey material should be treated, as far as possible, like a patient specimen.
4. The survey material should be treated as potentially hazardous and standard laboratory safety precautions should be taken as usual in your laboratory.
5. To complete the survey, you should send back the stained slides together with appropriate controls, and the questionnaire for statistical analysis and compilation of results.
6. Do NOT forward any particulars of your laboratory except your assigned confidential Laboratory Code. **Late return will be counted as not done and the results will NOT be accepted for assessment and statistical analysis.**
7. All stained slides should be properly labelled with the **assigned 4-digit laboratory code but not the identity of your laboratory.**
8. Place the labelled slides inside the slide mailers. Return the slide mailers and completed questionnaire in the enclosed foam-padded envelope.
9. Arrange return of survey material with the courier service provider according to the instruction as per attached before on **23 May 2024**

For further enquiry, please contact HKIMLSQAP Ltd.

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CONFIDENTIALITY

HKIMLSQAP is committed to keep all details of participants confidential. Please refer to <http://www.hkimlsqap.org>

**HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES
QUALITY ASSURANCE PROGRAMME
ANATOMICAL PATHOLOGY**

MAY SURVEY (2024)

HISTOLOGICAL STAINING MODULE

You are provided with two paraffin sections, which had been fixed in 10% buffered formalin. Perform the following staining methods and return the **STAINED SECTIONS** for assessment.

HC2406 Stain with your routine H&E method

HC2407 Stain with Periodic Acid Silver Methenamine (PASM) method to demonstrate the basement membrane of glomerulus in kidney

POSITIVE CONTROLS ARE REQUIRED BUT NOT FOR ASSESSMENT.

Please label the slides with the corresponding **Test code** and your confidential **Laboratory Code** for identification purposes. **No mark will be given for the unlabelled slide.** Put not more than two slides in one mailer in each return.



**HISTOLOGICAL STAINING MODULE
RETURN FORM**

Laboratory Code:

Date of Return: on or before **23 May 2024**

QUESTIONNAIRE

SECTION I: HISTOPATHOLOGY

1. What is the section thickness for your routine PASM stain in renal biopsy?

- 1 µm _____
- 2 µm _____
- 3 µm _____
- 4 µm _____
- Others (please state) _____

2. Do you use PASM method for demonstrating basement membranes of glomerulus in kidney?

- Yes _____
- No _____
- If NO, please specify the method _____

3. Do you freshly prepare the working PASM silver working solution?

- Yes _____
- No _____

4. Do you use commercial PASM silver working solution for demonstrating basement membranes of glomerulus in kidney?

- Yes _____
- No _____
- If YES, please specify the manufacturer _____

5. Do you filter the working PASM silver working solution before use

- Yes _____
- No _____
- Others (please state) _____

6. What type(s) of oxidizing agent(s) you used in the Oxidation step of PASM method?

Periodic acid _____
 Chromic acid _____
 Others (please state) _____

7. How long will the slide(s) be incubated with PASM silver working solution in water bath?

10 – 15 minutes _____
 15 – 30 minutes _____
 30 – 45 minutes _____
 Others (please state) _____

8. What temperature of water bath is set for incubation of PASM silver working solution?

9. What counterstain is used in your laboratory for PASM method?

PASM Method

<i>Reagent</i>	<i>% / Composition</i>	<i>Time</i>	<i>Temp (°C)</i>
Oxidizing agent(s)	_____ % _____	min.	
	_____ % _____	min.	
PASM silver working solution Home-made _____ Commercial _____	_____ ml _____ % _____	min.	
	_____ ml _____ % _____		
	_____ ml _____ % _____		
	_____ ml distilled water		
Toning solution	_____ % _____	min.	
Hypo	_____ % _____	min.	
Counterstain (if any)	_____ % _____	min.	
Other (if any)	_____ % _____	min.	

END

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ANATOMICAL PATHOLOGY**

MAY SURVEY (2024)

IMMUNOHISTOCHEMICAL STAINING MODULE

Evaluation of CD117 Antibody

HC2408 You are provided with a paraffin section, stain the slide with your **in-house Anti-Human CD117** antibody and your routine immunohistochemistry detection system.

Evaluation of Ki67 Demonstration

HC2409 You are provided with a paraffin section, stain the slide with your **in-house Anti-Human Ki67** antibody and your routine immunohistochemistry detection system.

Evaluation of CDX2 Demonstration

HC2410 You are provided with a paraffin section, stain the slide with your **in-house Anti-Human CDX2** antibody and your routine immunohistochemistry detection system.

POSITIVE CONTROLS ARE REQUIRED BUT NOT FOR ASSESSMENT.

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HONG KONG INSTITUTE OF MEDICAL LABORATORY SCIENCES
 QUALITY ASSURANCE PROGRAMME

IMMUNOHISTOCHEMICAL STAINING MODULE
 RETURN FORM

Laboratory Code:

Date of Return: on or before **23 May 2024**

QUESTIONNAIRE

Your immunohistochemistry conditions:

<i>STEP</i>	<i>HC2408 (CD117 in-house)</i>	<i>HC2409 (Ki67 in-house)</i>	<i>HC2410 (CDX2 in-house)</i>
Supplier			
Dilution	1:	1:	1:
Peroxidase Blocking	min.	min.	min.
Antigen retrieval:	YES / NO	YES / NO	YES / NO
<input type="checkbox"/> Trypsinization <input type="checkbox"/> Microwave <input type="checkbox"/> Pressure cooking <input type="checkbox"/> PT module <input type="checkbox"/> Others	min.	min.	min.
Detection System			
Duration of Colour Development	DAB min.	DAB min.	DAB min.
End product Colour Enhancement (if any)	min.	min.	min.

END